

# BAJet® DNA In Vitro Transfection Reagent (Ver. II)

---A General Protocol for transfecting Mammalian Cells

- ☐ 100 µl
- ☐ 500 µl
- ☐ 1000 µl



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This product is for laboratory research ONLY and not for diagnostic use

## Introduction:

BAJet® (Ver. II) is an enhanced liposome-based DNA transfection reagent which is specifically formulated and optimized for mammalian cells with superior efficiency and less cytotoxicity. BAJet® Reagent (Ver. II), 1.0 ml, is sufficient for 600 to 1200 transfections in 24 well plates or 300 to 600 transfections in 6 well plates.

## Important Guidelines for Transfection:

- BAJet® reagent was formulated for DNA transfection ONLY! The following standard protocols are for transfecting mammalian cells. The protocols for lentivirus production and insect cell transfection can be downloaded from our website
- For better efficiency, choosing a correct protocol is essential. We strongly encourage to use "General Protocol" first. If the "General Protocol" fails to give satisfactory result (e.g., less than 10%), try the "Advanced Protocol" in the back page
- For high efficiency and lower toxicity, transfect cells at high density. 70~80% confluency is highly recommended
- To lower cytotoxicity, transfect cells in presence of serum (10%) and antibiotics

## Part I. General Procedures for Transfecting Mammalian Cells

### Step I. Cell Seeding:

Cells should be plated 18 to 24 hours prior to transfection so that the monolayer cell density reaches to the optimal 70~80% confluency at the time of transfection. Complete culture medium with serum and antibiotics is freshly added to each well 30~60 minutes before transfection.

**Note:** For high efficiency and lower toxicity, high cell density (~80% confluency) is essential. High serum levels (>5%) usually do not have inhibitory effect on transfection efficiency.

**Table 1. Recommended Amounts for Different Culture Vessel Formats**

Culture Dish	Culture Medium (ml)	Plasmid DNA (µg)	Diluent Volume (ml)	BAJet® Reagent (µl)
48 well plate	0.3	0.25	2 x 0.015	0.75
12 well plate	0.75	0.75	2 x 0.038	2.25
6-well/35 mm dish	1.0	1	2 x 0.05	3.0
60 mm dish	2.8	2.5	2 x 0.10	7.5
10 cm dish	5.0	5	2 x 0.25	15
T75 flask	8.0	9 - 18	2 x 0.40	27 - 54
250 ml flask	18	25 - 35	2 x 0.8	75 - 105

## Step II. Preparation of BAJet®-DNA Complex and Transfection Procedures

**For different cell types, the optimal ratio of BAJet® (µL):DNA (µg) is around 3:1. We recommend the BAJet® (µL):DNA (µg) ratio of 3:1 as a starting point which usually gives satisfactory transfection efficiency with invisible cytotoxicity. To ensure the optimal size of complex particles, we recommend using serum-free DMEM with High Glucose to dilute DNA and BAJet® Reagent.**

The following protocol is given for transfection in 24-well plates, refer to **Table 1** for transfection in other culture formats. The optimal transfection conditions for a majority of adherent cell lines are given in the standard protocol described below.

- For each well, add 0.5 ml of complete medium with serum and antibiotics freshly 30~60 minutes before transfection.
- For each well, dilute 0.5 µg of DNA into 25 µl of serum-free DMEM with High Glucose. Gently pipette up and down 3~4 times to mix.
- For each well, dilute 1.5 µl of BAJet® reagent into 25 µl of serum-free DMEM with High Glucose. Gently pipette up and down 3~4 times to mix.

**Note:** Never use Opti-MEM to dilute BAJet® reagent and DNA, it will disrupt transfection complex.

- Add the diluted BAJet® Reagent **immediately** to the diluted DNA solution all at once. (**Important: do not mix the solutions in the reverse order !**)
- Immediately pipette up and down 3~4 times or vortex briefly to mix followed by incubation for ~10 minutes at room temperature to allow DNA-BAJet® transfection complexes to form.

**Note:** Never keep the DNA/BAJet® complex longer than 20 minutes

- Add the 50 µl BAJet®/ DNA mixture drop-wise onto the medium in each well and homogenize the mixture by gently swirling the plate.
- Remove BAJet®/DNA complex-containing medium and replace with complete serum/antibiotics containing medium 12~18 hours post transfection. **For sensitive cells, to lower cytotoxicity, remove BAJet®-DNA complex and replace with complete medium 5 hours after transfection.**
- Check transfection efficiency 24 to 48 hours post transfection.

# BAJet® DNA In Vitro Transfection Reagent (Ver. II)

--An Advanced Protocol for Transfecting Hard-to-transfect Mammalian Cells

- ☐ 100  $\mu$ l  
☐ 500  $\mu$ l  
☐ 1000  $\mu$ l



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## Part II. Advanced Protocol for Transfecting Hard-To-Transfect Mammalian Cells

**Important:** The advanced protocol for hard-to-transfect cells is provided only if general protocol fails to give satisfactory efficiency (e.g., less than 10% efficiency). **For transfecting primary cells which cannot be trypsinized, go directly to Step II, skip trypsinization and incubate freshly prepared primary cell pellet with transfection complex.**

### Step I. Cell Culture Before Transfection

Cells should be plated at least 24 hours prior to transfection so that the monolayer cell density reaches to the optimal 95~100% confluency at the day of transfection.

**Table 2. A Guideline for Optimal Cell Number Per Well in Different Culture Formats**

Culture Dishes	Surface Area (cm <sup>2</sup> )	Optimal Cell Number
T75 Flask	75	$9.6 \times 10^6$
100 mm Dish	58	$7.3 \times 10^6$
60 mm Dish	21	$2.7 \times 10^6$
35 mm Dish	9.6	$1.2 \times 10^6$
6-well Plate	9.6	$1.2 \times 10^6$
12-well Plate	3.5	$0.44 \times 10^6$
24-well Plate	1.9	$0.24 \times 10^6$
48-well Plate	1.0	$0.11 \times 10^6$
96-well Plate	0.3	$0.31 \times 10^5$

**Table 3. Recommended Amounts for Different Culture Vessel Formats**

Culture Dish	Transfection Complex Volume (ml)	Plasmid DNA ( $\mu$ g)	BAJet® Reagent ( $\mu$ L)
96-well	0.02	0.2	0.8
48-well	0.04	0.5	2
24-well	0.1	1.0	4
6-well	0.2	2	8
35 mm dish	0.2	2	8
60 mm dish	0.5	5	20
10 cm dish	1.0	8	32
T75 flask	1.5	36	144

### Step II. Preparation of Cells in Suspension

The following protocol is given for transfecting hard-to-transfect mammalian cells in 6-well plates, refer to **Table 2** for optimal cell

number per well per culture vessels' surface area. The optimal transfection conditions for mammalian cells are given in the standard protocol described below.

- Detach the cells with trypsin/EDTA and stop the trypsinization with complete culture medium.

**Note:** Cells that are difficult to detach may be placed at 37 °C for 5~15 min to facilitate detachment

- Take an aliquot of trypsinized cell suspension and count the cells to determine the cell density.
- Centrifuge the required  $1.2 \times 10^6$  cells per well for 6-well plate at 150xg at room temperature for 10 min.
- Use fine tip pipette to remove supernatant **completely** so that no residual medium covers the cell pellet.

### Step III. Preparation and Application of Transfection Complex

**For hard-to-transfect mammalian cells, the optimal ratio of BAJet® ( $\mu$ L):DNA ( $\mu$ g) is 4:1. To ensure the optimal size of complex particles, we recommend using serum-free DMEM with High Glucose to dilute DNA and BAJet® Reagent.**

The following protocol is given for transfection in 6-well plates, refer to **Table 3** for transfection in other culture formats.

- For each well of 6-well plate, dilute 2  $\mu$ g of DNA into 100  $\mu$ l of serum-free DMEM with High Glucose. Vortex gently and spin down briefly to bring drops to bottom of the tube.
- For each well of 6-well plate, dilute 8  $\mu$ l of BAJet® reagent (Ver. II) into 100  $\mu$ l of serum-free DMEM with High Glucose. Vortex gently and spin down briefly.
- Add the diluted BAJet® Reagent immediately to the diluted DNA solution all at once.
- Immediately pipette up and down 3~4 times or vortex briefly to mix followed by incubation for ~ 15 minutes at room temperature to allow DNA-BAJet® transfection complexes to form.
- **Gently** resuspend the cell pellet prepared from **Step II** immediately in the 200  $\mu$ l transfection complex and incubate at 37 °C for 15 minutes.

Important: Never keep the transfection complexes longer than 30 minutes

- At the end of incubation, add 2.0 ml of pre-warmed fresh complete cell growth medium to cells and plate onto one well of a 6-well plate.
- Remove transfection complex containing medium gently and refill with complete culture medium 12~18 hours after plating.
- Check efficiency 24~48 hours post transfection.